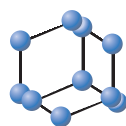


## RESEARCH ARTICLE

BENTHAM  
SCIENCE

# Age and Dietary Vitamin C Intake Affect Brain Physiology in Genetically Modified Mice Expressing Human Lipoprotein(A) and Unable to Synthesize Vitamin C

Lei Shi<sup>1</sup>, Aleksandra Niedzwiecki<sup>1,\*</sup> and Matthias Rath<sup>1</sup><sup>1</sup>Dr. Rath Research Institute 5941 Optical Ct, San Jose, CA 95138, USA

## ARTICLE HISTORY

Received: January 01, 2021  
 Revised: April 15, 2021  
 Accepted: May 09, 2021

DOI:  
 10.2174/1874609814666210706170326



CrossMark

**Abstract:** *Aims:* Lipoprotein (a) deposition in coronary vascular plaques and cerebral vessels is a recognized risk factor for cardiovascular disease, and research supports its role as a “repair factor” in vascular walls weakened by vitamin C deficiency.

**Background:** Humans depend on dietary vitamin C as an important antioxidant and as a cofactor in collagen synthesis, yet are prone to vitamin C deficiency. The brain is the one with the highest vitamin C content, owing to its high oxygen consumption and oxidative stress. It has been shown that brain aging is accompanied by accumulated oxidative damage, which can lead to memory decline and neurological diseases.

**Objective:** Our transgenic mouse, Gulo (-/-); Lp(a)<sup>+</sup>, presents a unique model for the study of key aspects of human metabolism with respect to a lack of internal vitamin C synthesis and the production of human lipoprotein(a).

**Methods:** This mouse model was used in our study to investigate the effects of prolonged intake of low and high levels of vitamin C, at different ages, on oxidative damage, cholesterol levels and lipoprotein(a) deposition in the brain.

**Result:** The results show that a long-term high vitamin C intake is important in maintaining brain cholesterol homeostasis and preventing oxidative damage in Gulo(-/-);Lp(a)<sup>+</sup> mice as they age. Moreover, we observed that the formation of brain lipoprotein(a) deposits was negatively correlated with brain level of vitamin C, thereby confirming its role as a stability factor for an impaired extracellular matrix.

**Conclusion:** Our study emphasizes the critical role of vitamin C in protecting brain health as we age.

**Other:** Our findings show that optimal vitamin C intake from early life to old age is important for brain health as it prevents oxidative stress damage and maintains cholesterol homeostasis in the brain. More importantly, the negative correlation between brain ascorbic levels and the formation of Lp(a) deposit on the choroid plexus further emphasizes the critical role of vitamin C in protecting brain health throughout the normal aging process.

**Keywords:** Vitamin C, human lipoprotein(a), brain physiology, transgenic mice, Gulo (-/-);Lp(a)<sup>+</sup>, antioxidant.

## 1. INTRODUCTION

Age-associated accumulation of oxidative damage is a contributing factor to memory decline and increased risk of neurological diseases [1, 2]. Vitamin C deficiency, which is implicated in reduced antioxidant defense in the brain, is related to increased risk of cognitive problems and cerebral diseases as well as accelerated amyloid accumulation in the

course of normal aging [3-6]. Higher vitamin C intake in elderly individuals is associated with reduced cognitive decline and a lower risk of Alzheimer's disease [7-9].

Vitamin C is not only an important antioxidant but it also regulates collagen production and serves as a cofactor in the hydroxylation of proline and lysine to form a triple helix during collagen biosynthesis [10]. This makes vitamin C essential in maintaining the stability of connective tissue and the integrity of the vascular walls.

Surprisingly, vitamin C deficiency has been frequently reported in Western populations [11-13]. An estimated 25%

\*Address correspondence to this author at Dr. Rath Research Institute 5941 Optical Ct, San Jose, CA 95138, USA. Email: [author@drrath.com](mailto:author@drrath.com)

of men and 16% of women in the low-income population in the UK had plasma vitamin C levels lower than 11 μmol/L, which is characteristic of scurvy [14]. A study in Canadian men and women aged 20-29 years showed that about 1 in 7 had vitamin C deficiency [15]. Aging is also associated with a risk of vitamin C deficiency owing to eating habits, accompanying diseases, and physiological changes [16].

Human dependency on dietary vitamin C intake stems from an inability to synthesize vitamin C endogenously, owing to the loss of the gene encoding for gluconolactone oxidase (Gulo), a key enzyme in the ascorbic acid biosynthesis pathway. This genetic change, which occurred in our primate ancestors about 60 million years ago, coincided with the emergence of Lp(a).

Lp(a) is a low-density cholesterol (LDL)-like lipoprotein, which contains a large protein, apolipoprotein(a) (apo(a)), linked by a disulfide bridge to the LDL protein component, apolipoprotein B-100 (apo(B)). The presence of Lp(a) is characteristic of human metabolism, and has been detected only in a very few animal species within a subset of primates [17]. Rath and Pauling suggested that owing to the presence of a large adhesive apo(a) protein, the Lp(a) can function as a repair molecule in the vascular walls, compensating for extracellular matrix (ECM) weakness, most often triggered by vitamin C deficiency [18, 19]. As this biological repair process overshoots, the large amount of lipids carried by this lipoprotein can turn this initial vascular repair process into atherosclerotic plaques [19]. We have shown earlier that loss of vascular endothelial integrity (increased endothelial gaps), observed during vitamin C deficiency, results in increased Lp(a) binding and the formation of atherosclerotic deposits in the vascular wall [20, 21].

Most studies of Lp(a) relate to coronary atherosclerosis. However, a study of random autopsies indicated that deposition of apo(a) was positively correlated with the degree of atherosclerosis detected in cerebral blood vessels [22]. Lp(a) has also been considered a risk factor for cerebrovascular disease and Alzheimer's disease [23, 24]. While apo(a) transcripts have been found in the brain and other organs [25], circulating apo(a) is synthesized mainly by the liver and secreted into the bloodstream [26]. There, it binds to circulating LDL to generate complete Lp(a) particles [27]. High Lp(a) concentration was found in patients with vascular dementia with cerebral infarction resulting from large artery occlusion [28]. Some studies suggested that abnormally high levels of serum Lp(a) are associated with cerebrovascular disease [29] and vascular dementia [30].

The brain contains approximately 25% of the total body cholesterol [31], which plays an essential role in brain development and function, including plasma membrane formation and synapse development. Numerous studies have shown that many pathological changes in the brain are associated with changes in brain cholesterol metabolism [32, 33]. It has been established that oxidative stress negatively impacts central nervous system (CNS) functions and is involved in neurodegenerative disorders, such as Alzheimer's disease [34]. Dixit *et al.* found that chronic vitamin C deficiency not only accelerated oxidative stress in mouse brains but also increased amyloid plaque production and cognitive impairment [35]. It has been suggested that high doses of vitamin C

could be protective against Alzheimer's disease-like pathologies [36]. Thus, the optimal intake of vitamin C is necessary to protect the brain from age-related oxidative stress as well as cerebrovascular disease.

To our knowledge, the information on the effects of chronic vitamin C deficiency in relation to the presence of Lp(a) and cholesterol metabolism in the brain during normal aging is scarce, mainly due to a lack of suitable animal models that can mimic human metabolism in respect of the Lp(a), coupled with a lack of vitamin C synthesis.

We developed a unique humanized mouse model unable to synthesize vitamin C (Gulo (-/-)) and at the same time carrying two mutations resulting in the synthesis of human apo(a) (h-apo(a)) and human apo(B)-100 (h-apo(B)-100) to form human Lp(a). This Gulo (-/-); Lp(a)+ mouse model has shown dietary vitamin C-related changes in collagen synthesis, Lp(a) production and atherogenicity, and cancer metastasis mediated through connective tissue stability [37].

In this study, we used the Gulo (-/-); Lp(a)+ mouse model to investigate the effects of the extended intake of low and high dietary vitamin C on age-related changes with respect to Lp(a) deposition in the brain, total brain cholesterol levels, and oxidative stress.

## 2. MATERIALS AND METHODS

### 2.1. Breeding and Genotyping of Transgenic Mice

Human Gulo (-/-); Lp(a)+ mice were generated as described by Cha *et al.* [20]. Briefly, homozygous Gulo (-/-) mice were generated by breeding heterozygous Gulo (+/-) mice BALB/cBy-Gulosfx/J (Jackson Laboratory, Sacramento, CA). Then homozygous Gulo (-/-) mice were bred with human apo(a) [h-apo(a)] transgenic mice (Mutant Mouse Regional Resource Center, Columbia, MO) and human apoB-100 (h-apoB-100) transgenic mice (Taconic Farms Inc., Hudson, NY) separately to produce Gulo(-/-); h-apo(a)+ mice and Gulo(-/-); h-apoB-100+ mice. These two transgenic mice were then bred to generate the Gulo (-/-); Lp(a)+ strain.

Genotyping was performed by TaqMan FAM probe Real Time-PCR at Transnetyx (Cordova, TN) using mouse tail clips. Genotyping ensured homozygosity of the Gulo locus knockout and the presence of h-apo (a) and h-apo(B)-100 genes. All animal experiments were conducted with humane and customary care, and followed a protocol approved by the internal Institutional Animal Care and Use Committee. All mice were housed in a barrier facility with a 12-hour light/12-hour dark cycle.

### 2.2. Experimental Design

Experiments were undertaken on both male and female Gulo (-/-); Lp(a)+ mice. Three groups of animals were used, aged 8-9 months (32-36 weeks), 1 year (52 weeks) and 2 years (104-116 weeks), at the time of harvesting. Twelve animals of each gender were randomly assigned to each age group. In each age group, 6 animals were assigned to a high vitamin C (H-VC)-supplemented diet, a modified LabDiet<sup>®</sup> Laboratory Rodent Diet 5001 with 1000 PPM vitamin C with distilled water, and 6 animals were assigned to a vitamin C-deficient (L-VC) diet (LabDiet<sup>®</sup> Laboratory Rodent

Diet 5001) with 30mg/L vitamin C added in distilled water. The H-VC diet provided the mice with approximately 4mg ascorbic acid daily, while the L-VC diet, with 30mg/L vitamin C added in distilled water, provided the mice with approximately 0.12mg ascorbic acid daily. Diet and water were provided *ad libitum*. The only source of vitamin C intake for the experimental Gulo (-/-);Lp(a)+ mice was the diet and water. We did not observe significantly different diet consumptions during the 20 weeks of the experiment. After 20 weeks, the mice were harvested for blood and tissues.

### 2.3. Sample Preparation

Serum was collected from blood drawn *via* cardiac puncture at the end of the experiment. Mouse midbrains were cut to preserve the hippocampus and were preserved in 10% neutral buffered formalin. Forebrain, hindbrain, and liver were collected, fast frozen in liquid nitrogen, and stored at -80°C until use.

### 2.4. Ascorbic Acid Measurement

Frozen mouse liver and brain were weighed and homogenized in Millipore water. The homogenates were then centrifuged at 2000rpm at 4 °C for 20 mins. The serum samples and tissue supernatants were used for ascorbic acid determination by a BioVision Ferric Reducing Ascorbate (FRASC) Assay Kit (Milpitas, CA), and expressed as nmole/mL for serum samples and nmole/mg tissue weight for brain samples.

### 2.5. Serum Apolipoproteins Measurement

Serum h-apo(a) levels were determined by using the IBL International GmbH Lp(a) Enzyme Immunoassay (Hamburg, Germany). Serum h-apoB-100 was determined by using the Assaypro AssayMax Human Apolipoprotein B Enzyme Kit (St. Charles, MO).

### 2.6. Brain Cholesterol Measurement

Brain total cholesterol levels were determined by Abcam HDL and LDL/VLDL Cholesterol Assay Elisa Kit (Cambridge, MA).

### 2.7. Brain 8-OHdG

Brain 8-OHdG levels were determined by BioVision QuickDetect™ 8-OHdG (Mouse) Elisa Kit (Milpitas, CA).

### 2.8. Immunohistology

Mouse midbrains were fixed in 10% neutral buffered formalin. The midbrains were then embedded in paraffin, sectioned and stained for hematoxylin and eosin (HE), h-apo(a), and h-apo(B)-100 at Histotox Labs, Inc. (Boulder, CO).

Immunohistochemical analysis was performed using ImageScope. The regions of interest (ROI) were the hippocampus region (1000 X 2800 microns) and the choroid plexus region (45X45 microns). Deposition of Lp(a) was indicated by the collocation of h-apo(a) and h-apo(B)-100. The extent of positive stains of h-apo(a) and h-apo(B)-100 was meas-

ured in the ROI for 3 mice per group. The data were expressed as the ratio of the number of positive pixels (positive stains) over the total number of pixels (positive+negative). The deposition of Lp(a) was calculated as the percentage of positive h-apo(a) stain over positive h-apo(B) stain at the same ROI.

### 2.9. Statistical Analysis

All data are presented as means ± standard deviation. Significant differences between means were determined by student's t-tests at a significance level of 0.05 with Microsoft Excel. The student's t-tests were computed between groups differing by a single characteristic. Specifically, t-tests were computed for each biomarker between H-VC and L-VC groups of the same age and sex, between age groups of the same vitamin C diet and gender, and between genders of the same vitamin C diet and age. Semi-partial correlation analysis between Lp(a) deposition rate in choroid plexus and brain vitamin C levels was performed by R package ppcor (Kim S., 2015).

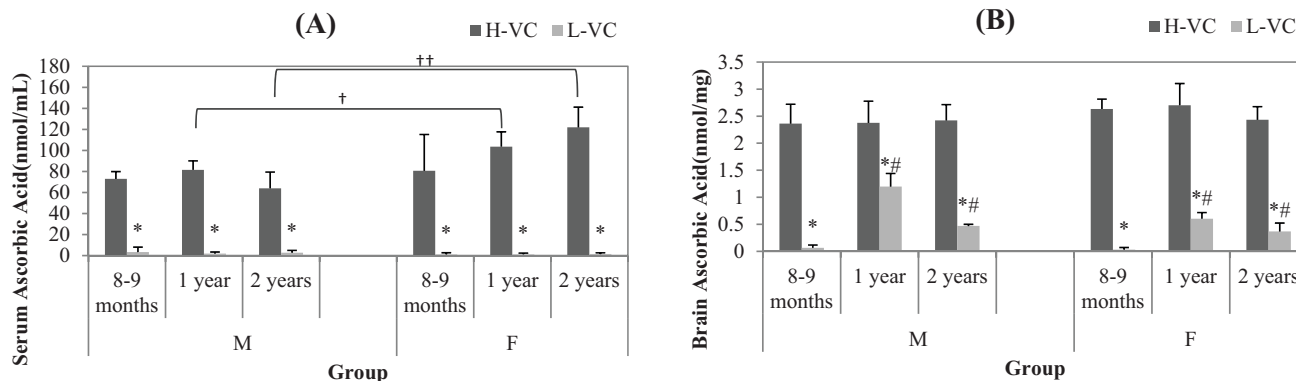
## 3. RESULTS

### 3.1. Serum Ascorbic Acid Levels

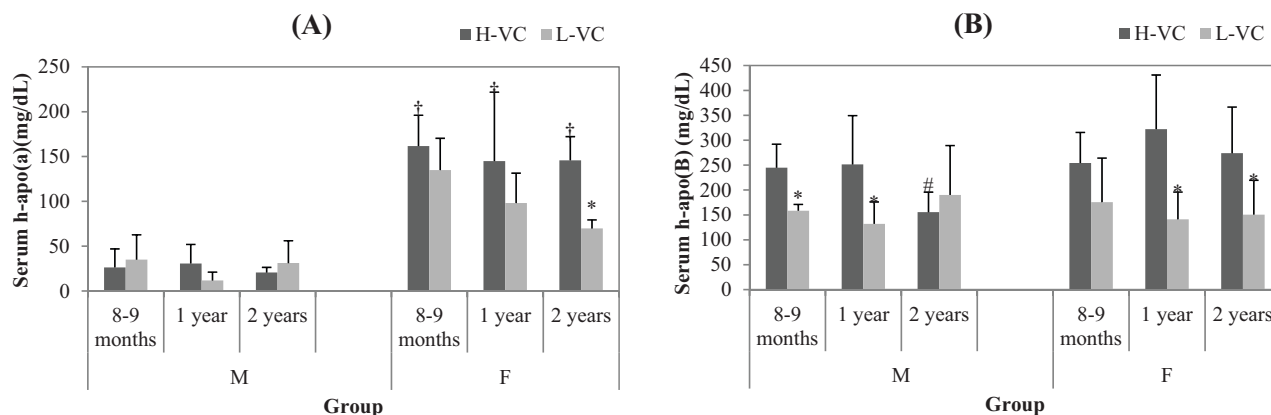
Age-related changes in ascorbic acid levels in the serum and brain tissue of Gulo (-/-); Lp(a)+ mice kept on diets supplemented with a high dose of vitamin C (H-VC) and a minimal dose of vitamin-C (L-VC) for 20 weeks are shown in Fig. (1).

Fig. (1A) shows that serum ascorbic acid levels were significantly lower in both male and female L-VC mice compared with mice consuming an H-VC diet ( $p < 0.05$ ). As such, serum ascorbic acid levels in L-VC males at the ages of 8-9 month, 1 year, and 2 years were 3.3, 1.8, and 2.7nmol/ml, respectively, and in L-VC females at the equivalent ages were 1.1, 1.2, and 1.5nmol/ml, respectively. Serum ascorbic acid levels in H-VC mice of corresponding sex and age were for males: 72.9, 81.5, and 63.9nmol/ml, respectively; while for females: 80.6, 103.6, and 121.9nmol/ml, respectively. Although serum ascorbic acid levels appear to increase with age in H-VC female mice, the differences did not reach statistical significance ( $p > 0.05$ ). However, female mice at the ages of 1 year and 2 years had significantly higher serum ascorbic acid levels than male mice of corresponding ages ( $p < 0.05$ ).

As shown in Fig. (1B), the brain ascorbic acid levels were several times lower in both male and female L-VC mice at the age of 8-9 months, 1 year and 2 years (male: 0.1, 1.2, and 0.5nmol/mg, respectively; female: 0.03, 0.6, and 0.4nmol/mg, respectively;  $p < 0.05$ ) compared with H-VC mice in corresponding sex and age groups (male: 2.4, 2.4, and 2.4nmol/mg, respectively; female: 2.6, 2.7, and 2.4nmol/mg, respectively). Whereas we had seen age-related changes in serum ascorbic acid levels, the H-VC mice maintained similar brain ascorbic acid levels across different age groups in both males and females. Interestingly, 1- and 2-year-old L-VC mice had significantly higher ascorbate contents in brain tissue than 8-9-month-old L-VC mice of both genders ( $p < 0.05$ ).



**Fig. (1).** Ascorbic acid levels in serum (A) and brain (B) in each age group and gender. Data are expressed as Mean  $\pm$  SD. \* represents statistically significant difference between H-VC groups and L-VC groups of the same age and gender at the significance level of 0.05; # represents statistically significant difference between age groups of the same gender and diet at the significance level of 0.05; † represents statistically significant difference between genders of the same age and diet at the significance level of 0.05; †† indicates  $p < 0.01$ . (A higher resolution / colour version of this figure is available in the electronic copy of the article).



**Fig. (2).** Serum levels of h-apo(a) (A) and h-apo(B) (B) in each age group and gender. Data are expressed as Mean  $\pm$  SD. \* represents statistically significant difference between H-VC groups and L-VC groups of the same age and gender at the significance level of 0.05; # represents statistically significant difference among age groups of the same gender and diet at the significance level of 0.05; † represents statistically significant difference between genders of the same age and diet at the significance level of 0.05. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

### 3.2. Serum Lipoproteins

#### 3.2.1. Serum Human Apo(a) Levels

As seen in Fig. (2A), serum h-apo(a) levels were significantly higher in female than in male mice of comparable age and dietary vitamin C intake. In L-VC female mice, serum h-apo(a) levels were 134.9, 98.1, and 69.7 mg/dL at the age of 8-9 months, 1 year and 2 years, respectively. Serum h-apo(a) levels in H-VC female mice at corresponding ages were higher, at 161.6, 144.9, and 145.7 mg/dL, respectively. However, the statistically significant difference in serum h-apo(a) between the L-VC and H-VC mice was observed only in the 2-year-old female mice ( $p < 0.05$ ). The L-VC female mice had approximately 2-fold lower serum h-apo(a) levels compared with H-VC female mice at the age of 2 years ( $p < 0.05$ ). Serum h-apo(a) levels in male mice did not markedly differ between L-VC groups at the age of 8-9 months, 1 year and 2

years (34.9, 11.7, and 31.1 mg/dL, respectively;  $p > 0.05$ ) and H-VC groups at corresponding ages (26.3, 30.6, and 20.5 mg/dL, respectively).

Interestingly, in both H-VC and L-VC groups, we found that serum h-apo(a) levels were approximately 5-fold higher in female mice than in male mice in all age groups ( $p < 0.05$ ). Furthermore, serum h-apo(a) levels did not differ among ages in H-VC and L-VC groups in either gender ( $p > 0.05$ ).

#### 3.2.2. Serum H-apo(B) Levels

Fig. (2B) shows the values for serum h-apo(B) in Gulo (-/-); Lp(a)+ mice fed an H-VC or L-VC diet for 20 weeks.

Serum h-apo(B) levels in 2-year-old H-VC male mice were significantly lower compared with 8-9-month- and 1-year-old animals (155.5, 244.7, and 251.3 mg/dL, respectively;  $p < 0.05$ ). However, these differences did not reach statis-

tical significance in H-VC female mice at corresponding ages (273.9, 254.2, and 322.2mg/dL, respectively;  $p>0.05$ ). L-VC female mice had lower serum h-apo(B) levels at the age of 8-9 months, and significantly lower levels at the age of 1 year and 2 years, compared with H-VC animals (175.5, 141.1, and 150.4mg/dL, respectively;  $p<0.05$ ). L-VC male mice at the age of 8-9 months and 1 year had less h-apo(B) than H-VC animals of corresponding ages (158.3 and 131.9mg/dL, respectively;  $p<0.05$ ). However, 2-year-old L-VC male mice had slightly higher serum h-apo(B) (190.1mg/dL) than H-VC animals (155.5 mg/dL).

### 3.2.3. Brain Total Cholesterol

Fig. (3A) shows age-related changes in brain total cholesterol levels in L-VC and H-VC mice.

The brain total cholesterol levels in younger male and female H-VC mice were significantly higher than in 2-year-old animals. As such, males aged 8-9 months and 1 year had cholesterol levels of 3.4 and 3.9 ug/mg, respectively ( $p>0.05$ ), and females 2.6 and 4.2 ug/mg, respectively ( $p<0.01$ ). Brain total cholesterol level was significantly lower in 2-year-old mice of both genders (males: 1.9 ug/mg; and females: 2.5 ug/mg;  $p<0.01$ ).

Brain cholesterol levels in female mice were affected more by age than by vitamin C dietary intake. Brain cholesterol levels in H-VC mice did not significantly differ from those in L-VC mice at the age of 8-9 months and 1 year in either gender. As such, brain cholesterol levels in L-VC males were 2.9 and 4.1ug/mg, respectively; and in L-VC females were 2.9 and 4.7ug/mg, respectively ( $p>0.05$ ). However, the brain cholesterol level was significantly lower in L-VC mice than in H-VC mice at the age of 2 years (males: 1.3 and 1.9ug/mg; females: 1.6 and 2.5ug/mg, respectively;  $p<0.05$ ).

### 3.2.4. Brain Oxidation

To further evaluate the age-related metabolic changes in the brain of Gulo (-/-); Lp(a)+ mice with high and low dietary vitamin C intake, we examined an important biomarker for cellular oxidative stress - 8-hydroxy-2'-deoxyguanosine (8-OHdG).

Fig. (3B) shows that in H-VC groups, brain 8-OHdG levels gradually increased with age in both male and female mice. In H-VC male mice aged 8-9 months, 1 year and 2 years, brain 8-OHdG levels were 5.2, 5.5, and 6.2ng/mg, respectively; and in females were 5.3, 5.9, and 6.2ng/mg, respectively. Brain 8-OHdG levels were significantly higher in male mice aged 2 than in males aged 8-9-months ( $p<0.05$ ), but not in female mice ( $p=0.052$ ).

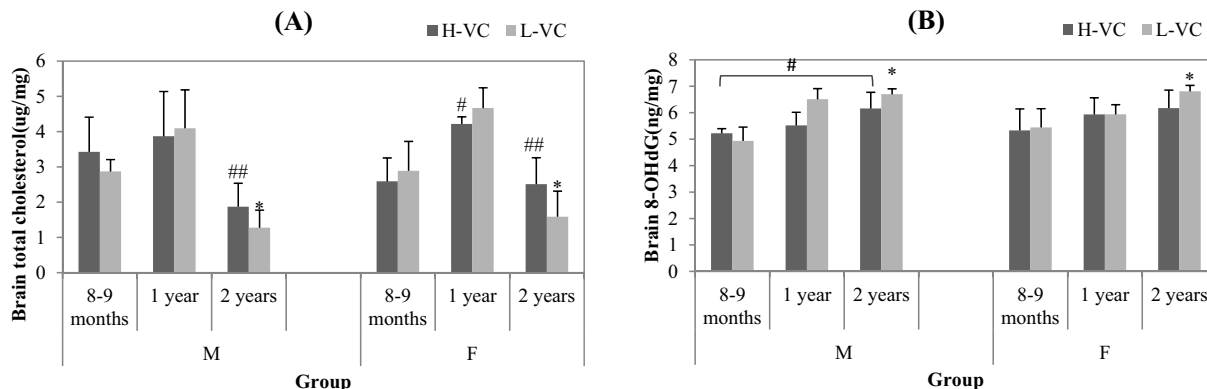
Compared with H-VC groups, brain 8-OHdG levels significantly increased in 2-year-old L-VC mice of both genders (male: 6.7ng/mg and female: 6.8ng/mg, respectively;  $p<0.05$ ). However, the differences in brain 8-OHdG levels were not statistically significant between mice on H-VC and L-VC diet at the age of 8-9 months and 1 year (male: 4.9 and 6.5ng/mg, respectively; and female: 5.4 and 5.9ng/mg, respectively;  $p>0.05$ ).

### 3.2.5. Brain H-apo(a) and H-apo(B) Deposition

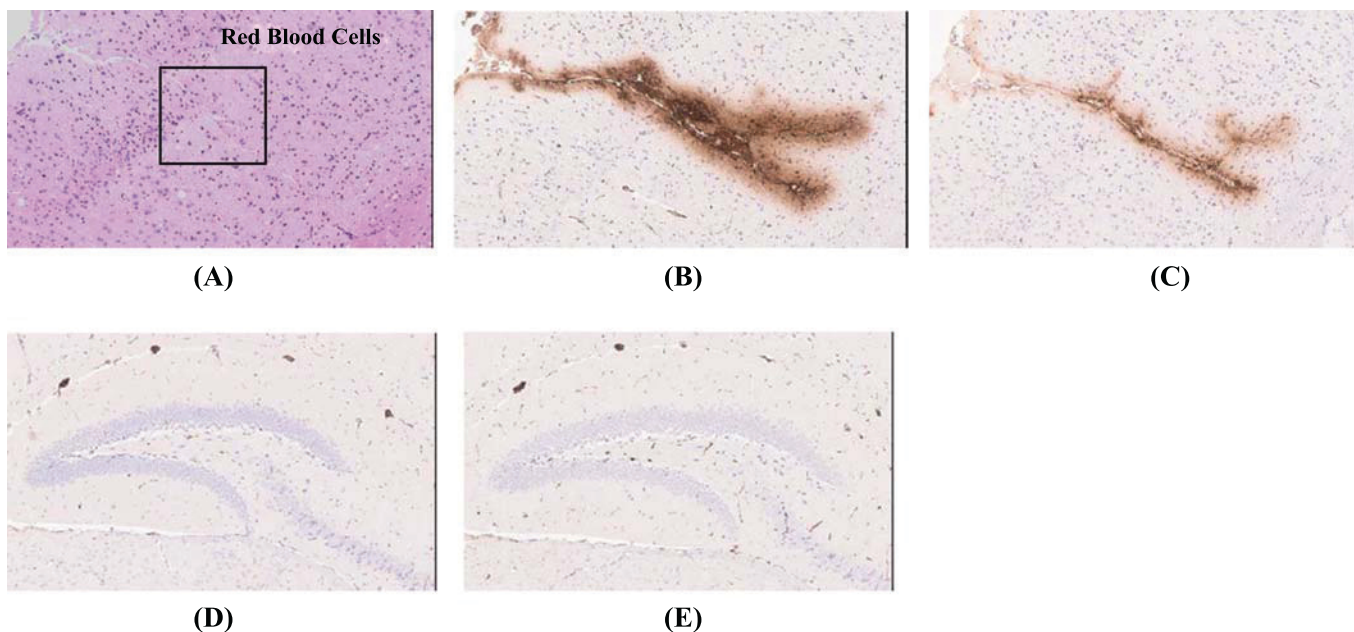
Mouse brains isolated from both H-VC and L-VC mice displayed areas of microhemorrhages. The microhemorrhages were mostly seen in cortical vessels, as shown in Fig. (4A). The collocations of h-apo(a) and h-apo(B)-100 immunostainings indicative of the Lp(a) deposition were found in brain capillaries, hemorrhaged blood vessels, and the choroid plexus, as shown in Figs. (4B-E and 5A-B).

The observed most intense h-apo(a) and h-apo(B)-100 immunoreactivity was related to hemorrhage in cortical vessels and brain capillaries, and to a lesser extent to non-leaky blood vessels. In addition, we observed mild mouse MMP9 stains co-located with h-apo(a) and h-apo(B)-100 deposition at the hemorrhaged blood vessels (picture not shown), indicating a possible injury or blood-brain dysfunction at these areas.

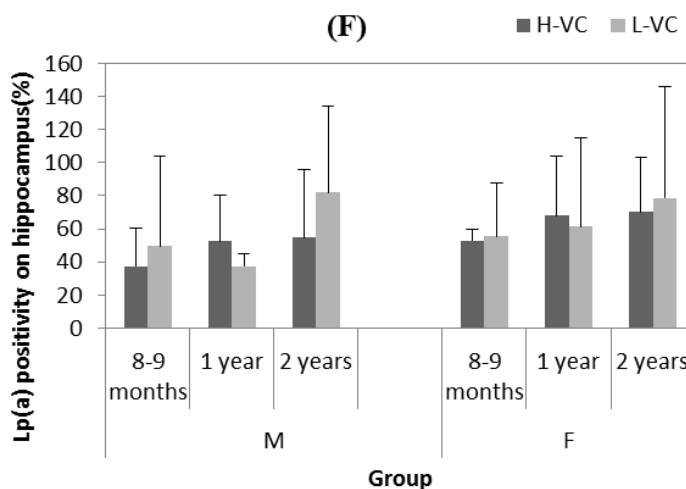
The degree of Lp(a) deposition in the hippocampus was expressed as the percentage of h-apo(a) and h-apo(B)-100 collocation (Fig. 4F). In H-VC mice, the Lp(a) deposits increased with age both in males and females. As such, the Lp(a) deposit rate increased in H-VC male mice from 37% at 8-9 months, to 53% in 1-year-old mice and 55% in 2-year-old mice. In H-VC female mice of corresponding ages, the Lp(a) tissue deposition increased from 53% to 68%, and to 70%, respectively.



**Fig. (3). Brain total cholesterol (A) and brain 8-OHdG (B) levels in each group of both genders.** Data are expressed as Mean  $\pm$  SD. \* represents statistically significant difference between H-VC groups and L-VC groups of the same age and gender at the significance level of 0.05; # represents statistically significant difference among age groups of the same gender and diet at the significance level of 0.05; ## indicates  $p<0.01$ . (A higher resolution / colour version of this figure is available in the electronic copy of the article).



**Fig. (4).** Examples of microhemorrhage HE staining (A), h-apo(a) deposit (B), and h-apo(B)-100 deposit (C) in brain cortical vessels. Examples of h-apo(a) deposits (D) and h-apo(B)-100 deposits (E) in the hippocampus region. Examples of HE stain. (A higher resolution / colour version of this figure is available in the electronic copy of the article).



**Fig. (4). (F) Quantification of Lp(a) deposit on the hippocampus** (A) The Lp(a) positivity was calculated as the percentage of h-apo(a) stain over h-apo(B)-100 stain at the same areas in the hippocampus. Data are expressed as Mean  $\pm$  SD. \* represents statistically significant difference between H-V C groups and L-VC- groups at the  $p$  level of 0.05; # represents statistically significant differences between age groups at the  $p$  level of 0.05. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

The differences in Lp(a) deposition in the hippocampus between H-VC and L-VC mice of the same age and diet did not reach statistical significance. We may have been underpowered to detect a difference. However, we could observe an evident trend towards higher Lp(a) deposition in L-VC mice, especially at the ages of 8-9 months and 2 years. As such, Lp(a) deposits in L-VC male mice at the ages of 8-9 months and 2 years were 50% and 82%, respectively, compared with 37% and 55% in male H-VC mice of correspond-

ing age ( $p > 0.05$ ). In L-VC females, the Lp(a) deposit rates in mice aged 8-9 months and 2 years were 56% and 79%, respectively, compared with 53% and 70% in H-VC females of corresponding ages, higher but not statistically different ( $p > 0.05$ ).

The degree of Lp(a) deposition on the choroid plexus was expressed as the percentage of h-apo(a) and h-apo(B)-100 immunostain collocation (Fig. 5B). The results showed increased deposition of Lp(a) in both male and female mice in

L-VC groups compared with H-VC animals at corresponding ages. In H-VC female mice, the Lp(a) deposits significantly increased from 63% to 90% and 93% in the 8-9 months, 1-year old and 2-year-old animals, respectively ( $p < 0.05$ ). However, the Lp(a) deposits in the choroid plexus in H-VC male mice of the corresponding ages were fairly consistent (54%, 53% and 58%, respectively;  $p > 0.05$ ).

Lp(a) deposits in the choroid plexus in L-VC mice were significantly higher at the age of 8-9 months, than in H-VC mice of corresponding age, in both genders (males: 118% and 54%, respectively; females: 133% and 63%, respectively;  $p < 0.05$ ). L-VC mice aged 2-years also had more Lp(a) deposits compared with H-VC mice of the same age (males: 134% and 58%, respectively; females: 129% and 93%, respectively;  $p > 0.05$ ).

The Lp(a) deposition was observed in the stromal space of the choroid plexus in both H-VC and L-VC mice. The representative micrographs of h-apo(a) and h-apo(b)-100 collocation in choroid plexus in each group are shown in Fig. (5A-B). The degree of Lp(a) deposition on the choroid plex-

us was expressed as the percentage of h-apo(a) and h-apo(b)-100 immunostain collocation (Fig. 5C). In H-VC female mice, the Lp(a) deposits increased from 63% to 82% in the 8-9 months and 2-year-old animals, respectively ( $p > 0.05$ ). The Lp(a) deposits in the choroid plexus in H-VC male mice of the corresponding ages were fairly consistent (54%, 53% and 58%, respectively;  $p > 0.05$ ).

Lp(a) deposits in the choroid plexus in L-VC mice were significantly higher at the age of 8-9 months, than in H-VC mice of corresponding age, in both genders (males: 118% and 54%, respectively; females: 133% and 63%, respectively;  $p < 0.05$ ). Female L-VC mice aged 1 year also showed more Lp(a) deposits compared with H-VC mice of the same age (132% and 65%, respectively;  $p > 0.05$ ).

Semi-partial regression analysis of values using individual animals, including both genders, all ages and vitamin C diets, revealed that brain ascorbic acid levels were significantly negatively correlated with Lp(a) deposition in choroid plexus ( $r^2 = 0.15$ ,  $p < 0.05$ , Fig. 5D).

(A)

Male mice: Lp(a) deposition in choroid plexus.

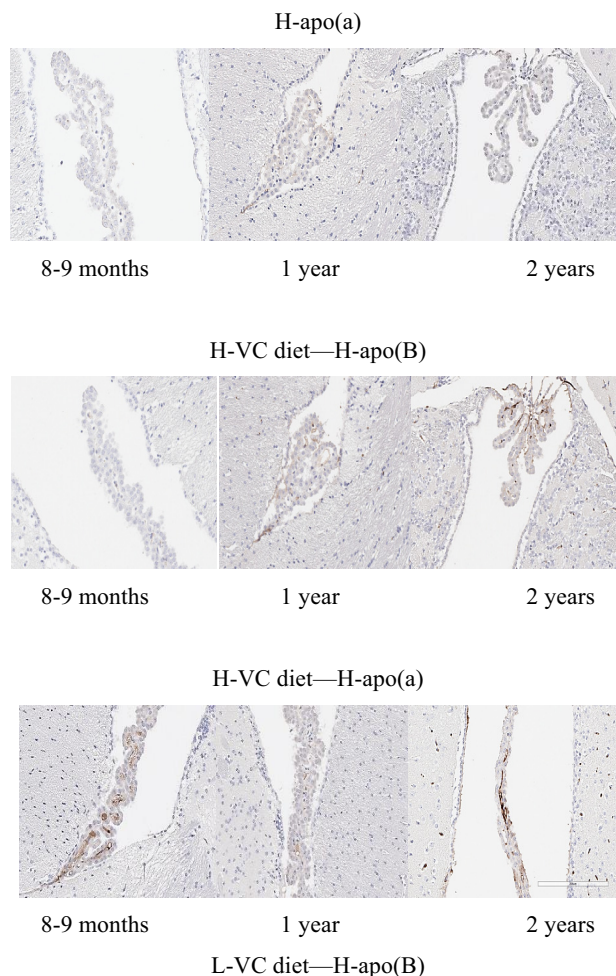
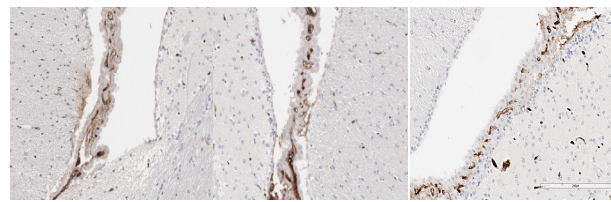


Fig. (5). Contd...



8-9 months

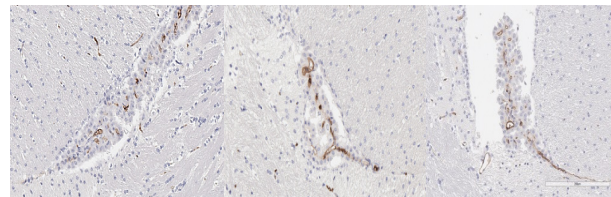
1 year

2 years

**(B)**

Female mice: Lp(a) deposition in choroid plexus.

H-apo(a)

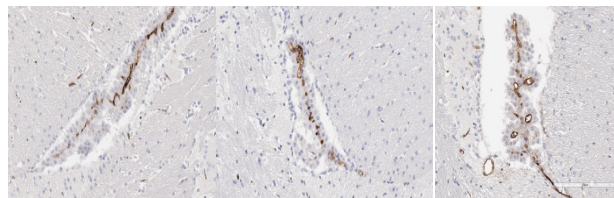


8-9 months

1 year

2 years

H-VC diet—H-apo(B)

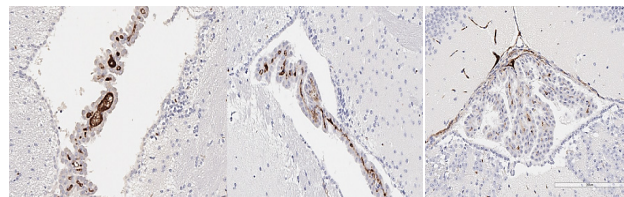


8-9 months

1 year

2 years

H-VC diet—H-apo(a)

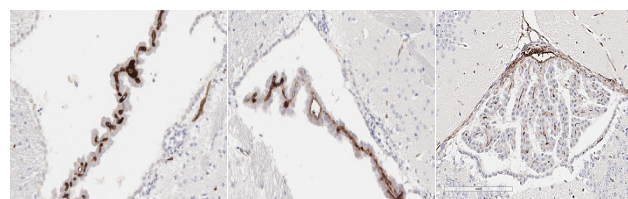


8-9 months

1 year

2 years

L-VC diet—H-apo(B)



8-9 months

1 year

2 years

**Fig. (5).** Contd...





Interestingly, older *Gulo* (-/-); *Lp(a)*<sup>+</sup> mice, especially at 1 year of age, kept on a low-ascorbate diet (L-VC), had significantly higher brain ascorbic acid levels than young 8-9-month-old mice. This corresponds to the findings that, in a state of vitamin C deprivation, the SCVT2 expression in the brain of aged animals is upregulated compared with young animals, which contributes to the increased retention of vitamin C in aged brains [43]. It is also possible that adult and aged mouse brains have greater buffering capacity, with regard to vitamin C retention, than the brains of young mice.

Brain cholesterol is synthesized *de novo* and its rate declines with age. Our results show that the brain cholesterol levels in *Gulo* (-/-); *Lp(a)*<sup>+</sup> mice were significantly lower in 2-year-old male and female mice compared with younger animals in both H-VC and L-VC groups. Vitamin C deficiency did not affect brain cholesterol levels in younger mice, which suggests that cholesterol metabolism in the brain of older mice is more prone to the consequences of scurvy. Although the role of brain cholesterol levels in neurological diseases is still unclear, it has been widely reported that Alzheimer's disease is accompanied by lower brain cholesterol levels [44, 45]. It has further been reported that brains with decreased levels of brain cholesterol are more susceptible to pathological diseases [46].

The brain is an organ with a high rate of oxygen consumption, thus it is prone to oxidative damage [47, 48], and numerous studies have reported an age-related increase of 8-OHdG in the brain [47, 49, 50]. Our results showed that 8-OHdG levels were significantly higher in the brains of 2-year-old *Gulo* (-/-); *Lp(a)*<sup>+</sup> mice compared with younger mice in both males and females, suggesting increased oxidative damage with age.

We found an increase of brain 8-OHdG levels in 2-year-old male mice and female mice kept for 20 weeks on a low vitamin C diet (L-VC), compared with mice consuming high levels of vitamin C (H-VC). Vitamin C consumption levels were not seen to affect 8-OHdG formation in the brain in younger mice in this study. Age-related changes were less pronounced than the changes observed at different vitamin C intake levels in both male and female mice. It is well established that vitamin C, as an antioxidant, plays an important role in protecting the brain from oxidative stress. When the brain is depleted of antioxidants, it becomes more susceptible to reactive oxygen species-related damage [51]. The accumulation of 8-OHdG in the brain is a contributing factor to neurodegenerative diseases such as Alzheimer's disease [52]. Our results suggest that great attention should be paid to avoiding vitamin C deficiency in mid to old-age to reduce the risk of impaired brain cholesterol levels and to protect against age-related oxidative stress in the brain.

Our results show that female mice had significantly higher serum h-apo(a) levels compared with male mice regardless of age and vitamin C intake. This gender difference was most likely owing to the suppressing effect of testosterone on h-apo(a) expression in male mice [53]. Such effect was supported by findings that the serum levels of h-apo(a) dramatically increase with castration in male h-apo(a) transgenic mice, but no consistent effect of castration was seen on serum levels of h-apo(a) in female h-apo(a) transgenic mice [54]. Human studies have indicated that estrogen can lower

*Lp(a)* levels [55], however, it was not observed in *Gulo*(-/-); *Lp(a)*<sup>+</sup> mice.

The majority of *Lp(a)* studies relate to its role as a risk factor for coronary heart disease. The cerebral arteries, as proximal arteries, are exposed to relatively high mechanical stress and are at a high risk of atherosclerosis-related diseases, especially in patients with low ascorbate levels [56]. However, the role of *Lp(a)* in brain aging and its related deposition in cerebral capillaries has not been previously elaborated. In this study, we found that *Lp(a)* was deposited in hemorrhaged cerebral capillaries and blood vessels, much as we observed in our previous studies showing *Lp(a)* deposition in the aorta and coronary blood vessels [20, 21]. This may reflect that both tissue repair and an anti-fibrinolytic mechanism as h-apo(a), made of repeated copies of Kringle IV plasminogen, can compete with plasminogen for its binding sites on endothelial cells, thus inhibiting fibrinolysis.

In addition, we found predominant *Lp(a)* deposition in the stromal side of the choroid plexus, which was significantly negatively correlated with the brain ascorbic acid levels. This finding corroborates our previous study results showing that *Lp(a)* accumulated in the vascular wall of dietary vitamin C-deficient *Gulo*(-/-); *Lp(a)* mice, but was absent in animals with high vitamin C intake [20], as well as is in accordance with *Lp(a)*'s role as a repair factor for a structurally weakened vascular wall [18]. The choroid plexus consists of many fenestrated capillaries lying on the basal membrane, containing type IV collagen. It has been shown that vitamin C deficiency lowers the expression of type IV collagen, thus affecting blood vessels integrity [57]. Therefore, the basal membrane of the choroid plexus in vitamin C-deficient mice could be damaged owing to chronic vitamin C deficiency. The choroid plexus functions as a barrier between blood and cerebrospinal fluid (CSF). It has been suggested that the cholesterol accumulation on the choroid plexus is one of the signs of choroid plexus damage [58].

## CONCLUSION

Our findings show that optimal vitamin C intake from early life to old age is important in brain health to prevent oxidative stress damage and to maintain cholesterol homeostasis in the brain. More importantly, the negative correlation between brain ascorbic levels and the formation of *Lp(a)* deposit on the choroid plexus further emphasizes the critical role of vitamin C in protecting brain health throughout the normal aging process.

## ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The study was approved by the internal Institutional Animal Care and Use Committee.

## HUMAN AND ANIMAL RIGHTS

The animal studies were ethically conducted, and the procedures were followed according to the research standards set by the National Academy of Sciences, The National Academies Press, Washington D.C.

**CONSENT FOR PUBLICATION**

Not applicable.

**AVAILABILITY OF DATA AND MATERIAL**

Not applicable.

**FUNDING**

Dr. Rath Health Foundation provided funding for this study. Grant no: RI-0-2020.

**CONFLICT OF INTEREST**

The authors declare no conflict of interest, financial or otherwise.

**ACKNOWLEDGEMENT**

The authors thank Dr. Ethan Jewett for the statistical evaluation. We thank Dr. Bilwa Bhanap for the help during the preparation and submission of this manuscript.

**REFERENCES**

- [1] Rodriguez KA, Wywiał E, Perez VI, *et al.* Walking the oxidative stress tightrope: A perspective from the naked mole-rat, the longest-living rodent. *Curr Pharm Des* 2011; 17(22): 2290-307. <http://dx.doi.org/10.2174/138161211797052457> PMID: 21736541
- [2] Harrison FE, May JM. Vitamin C function in the brain: Vital role of the ascorbate transporter SVCT2. *Free Radic Biol Med* 2009; 46(6): 719-30. <http://dx.doi.org/10.1016/j.freeradbiomed.2008.12.018> PMID: 19162177
- [3] Gale CR, Martyn CN, Winter PD, Cooper C. Vitamin C and risk of death from stroke and coronary heart disease in cohort of elderly people. *BMJ* 1995; 310(6994): 1563-6. <http://dx.doi.org/10.1136/bmj.310.6994.1563> PMID: 7787644
- [4] Gale CR, Martyn CN, Cooper C. Cognitive impairment and mortality in a cohort of elderly people. *BMJ* 1996; 312(7031): 608-11. <http://dx.doi.org/10.1136/bmj.312.7031.608> PMID: 8595334
- [5] Bulpitt CJ. Vitamin C and vascular disease. *BMJ* 1995; 310(6994): 1548-9. <http://dx.doi.org/10.1136/bmj.310.6994.1548> PMID: 7540454
- [6] Charlton KE, Rabinowitz TL, Geffen LN, Dhansay MA. Lowered plasma vitamin C, but not vitamin E, concentrations in dementia patients. *J Nutr Health Aging* 2004; 8(2): 99-107. PMID: 14978605
- [7] Wengreen HJ, Munger RG, Corcoran CD, *et al.* Antioxidant intake and cognitive function of elderly men and women: The Cache County Study. *J Nutr Health Aging* 2007; 11(3): 230-7. PMID: 17508099
- [8] Fotuhi M, Zandi PP, Hayden KM, *et al.* Better cognitive performance in elderly taking antioxidant vitamins E and C supplements in combination with nonsteroidal anti-inflammatory drugs: The Cache County Study. *Alzheimers Dement* 2008; 4(3): 223-7. <http://dx.doi.org/10.1016/j.jalz.2008.01.004> PMID: 18631971
- [9] Engelhart MJ, Geerlings MI, Ruitenberg A, *et al.* Dietary intake of antioxidants and risk of Alzheimer disease. *JAMA* 2002; 287(24): 3223-9. <http://dx.doi.org/10.1001/jama.287.24.3223> PMID: 12076218
- [10] Stetten MR. Some aspects of the metabolism of hydroxyproline, studied with the aid of isotopic nitrogen. *J Biol Chem* 1949; 181(1): 31-7. [http://dx.doi.org/10.1016/S0021-9258\(18\)56621-7](http://dx.doi.org/10.1016/S0021-9258(18)56621-7) PMID: 15390388
- [11] Fain O. Vitamin C deficiency. *Rev Med Interne* 2004; 25(12): 872-80. <http://dx.doi.org/10.1016/j.revmed.2004.03.009> PMID: 15582167
- [12] Hampl JS, Taylor CA, Johnston CS. Vitamin C deficiency and depletion in the United States: The third national health and nutrition examination survey, 1988 to 1994. *Am J Public Health* 2004; 94(5): 870-5. <http://dx.doi.org/10.2105/AJPH.94.5.870> PMID: 15117714
- [13] Schleicher RL, Carroll MD, Ford ES, Lacher DA. Serum vitamin C and the prevalence of vitamin C deficiency in the United States: 2003-2004 national health and nutrition examination survey (NHANES). *Am J Clin Nutr* 2009; 90(5): 1252-63. <http://dx.doi.org/10.3945/ajcn.2008.27016> PMID: 19675106
- [14] Mosdøl A, Erens B, Brunner EJ. Estimated prevalence and predictors of vitamin C deficiency within UK's low-income population. *J Public Health (Oxf)* 2008; 30(4): 456-60. <http://dx.doi.org/10.1093/pubmed/fdn076> PMID: 18812436
- [15] Cahill L, Corey PN, El-Soheby A. Vitamin C deficiency in a population of young Canadian adults. *Am J Epidemiol* 2009; 170(4): 464-71. <http://dx.doi.org/10.1093/aje/kwp156> PMID: 19596710
- [16] Zagaria MAE. Vitamin deficiencies in seniors. *US Pharm* 2010; 35(8): 20-7.
- [17] Tomlinson JE, McLean JW, Lawn RM. Rhesus monkey apolipoprotein(a). Sequence, evolution, and sites of synthesis. *J Biol Chem* 1989; 264(10): 5957-65. [http://dx.doi.org/10.1016/S0021-9258\(18\)83643-2](http://dx.doi.org/10.1016/S0021-9258(18)83643-2) PMID: 2925643
- [18] Rath M, Pauling L. Hypothesis: Lipoprotein(a) is a surrogate for ascorbate. *Proc Natl Acad Sci USA* 1990; 87(16): 6204-7. <http://dx.doi.org/10.1073/pnas.87.16.6204> PMID: 2143582
- [19] Rath M, Pauling L. Solution to the puzzle of human cardiovascular disease: Its primary cause is ascorbate deficiency leading to the deposition of lipoprotein(a) and fibrinogen/fibrin in the vascular wall. *J Orthomol Med* 1991; 6: 125-34.
- [20] Cha J, Niedzwiecki A, Rath M. Hypoascorbemia induces atherosclerosis and vascular deposition of lipoprotein(a) in transgenic mice. *Am J Cardiovasc Dis* 2015; 5(1): 53-62. PMID: 26064792
- [21] Shi L, Niedzwiecki A, Ivanov V, Rath M. Cardiovascular effects of cyclical dietary vitamin c withdrawal in mice deficient in internal synthesis vitamin c and producing human lipoprotein (a): Gulo(-/-); Lp(a)+. *Int J Cardiovasc Res* 2019; 8(1). <http://dx.doi.org/10.4172/2324-8602.1000369>
- [22] Jamieson DG, Usher DC, Rader DJ, Lavi E. Apolipoprotein(a) deposition in atherosclerotic plaques of cerebral vessels. A potential role for endothelial cells in lesion formation. *Am J Pathol* 1995; 147(6): 1567-74. PMID: 7495281
- [23] Iwamoto T, Watanabe D, Umahara T, Sakurai H, Hanyu H, Kanaya K. Dual inverse effects of lipoprotein(a) on the dementia process in Japanese late-onset Alzheimer's disease. *Psychogeriatrics* 2004; 4: 64-71. <http://dx.doi.org/10.1111/j.1479-8301.2004.00063.x>
- [24] Zenker G, Költringer P, Boné G, Niederkorn K, Pfeiffer K, Jürgens G. Lipoprotein(a) as a strong indicator for cerebrovascular disease. *Stroke* 1986; 17(5): 942-5. <http://dx.doi.org/10.1161/01.STR.17.5.942> PMID: 2945294
- [25] Ramharack R, Spahr MA, Kreick JS, Secker CS. Expression of apolipoprotein[a] and plasminogen mRNAs in cynomolgus monkey liver and extrahepatic tissues. *J Lipid Res* 1996; 37(9): 2029-40. [http://dx.doi.org/10.1016/S0022-2275\(20\)37567-2](http://dx.doi.org/10.1016/S0022-2275(20)37567-2) PMID: 8895068
- [26] Lobentanz EM, Krasznai K, Gruber A, *et al.* Intracellular metabolism of human apolipoprotein(a) in stably transfected Hep G2 cells. *Biochemistry* 1998; 37(16): 5417-25. <http://dx.doi.org/10.1021/bi972761t> PMID: 9548923
- [27] Kostner GM, Wo X, Frank S, Kostner K, Zimmermann R, Steyrer E. Metabolism of Lp(a): Assembly and excretion. *Clin Genet* 1997; 52(5): 347-54. <http://dx.doi.org/10.1111/j.1399-0004.1997.tb04352.x> PMID: 9520125
- [28] Urakami K, Mura T, Takahashi K. Lp(a) lipoprotein in cerebrovascular disease and dementia. *Jpn J Psychiatry Neurol* 1987; 41(4): 743-8. <http://dx.doi.org/10.1111/j.1440-1819.1987.tb00433.x> PMID: 2969061
- [29] Jürgens G, Taddei-Peters WC, Költringer P, *et al.* Lipoprotein(a) serum concentration and apolipoprotein(a) phenotype correlate

- with severity and presence of ischemic cerebrovascular disease. *Stroke* 1995; 26(10): 1841-8.  
<http://dx.doi.org/10.1161/01.STR.26.10.1841> PMID: 7570736
- [30] Urakami K, Wada-Isoe K, Wakutani Y, *et al.* Lipoprotein(a) phenotypes in patients with vascular dementia. *Dement Geriatr Cogn Disord* 2000; 11(3): 135-8.  
<http://dx.doi.org/10.1159/000017226> PMID: 10765043
- [31] Björkhem I, Meaney S. Brain cholesterol: Long secret life behind a barrier. *Arterioscler Thromb Vasc Biol* 2004; 24(5): 806-15.  
<http://dx.doi.org/10.1161/01.ATV.0000120374.59826.1b> PMID: 14764421
- [32] Puglielli L, Tanzi RE, Kovacs DM. Alzheimer's disease: The cholesterol connection. *Nat Neurosci* 2003; 6(4): 345-51.  
<http://dx.doi.org/10.1038/nn0403-345> PMID: 12658281
- [33] Ghribi O, Larsen B, Schrag M, Herman MM. High cholesterol content in neurons increases BACE, beta-amyloid, and phosphorylated tau levels in rabbit hippocampus. *Exp Neurol* 2006; 200(2): 460-7.  
<http://dx.doi.org/10.1016/j.expneurol.2006.03.019> PMID: 16696972
- [34] Salim S. Oxidative stress and the central nervous system. *J Pharmacol Exp Ther* 2017; 360(1): 201-5.  
<http://dx.doi.org/10.1124/jpet.116.237503> PMID: 27754930
- [35] Dixit S, Bernardo A, Walker JM, *et al.* Vitamin C deficiency in the brain impairs cognition, increases amyloid accumulation and deposition, and oxidative stress in APP/PSEN1 and normally aging mice. *ACS Chem Neurosci* 2015; 6(4): 570-81.  
<http://dx.doi.org/10.1021/cn500308h> PMID: 25642732
- [36] Kook SY, Lee KM, Kim Y, *et al.* High-dose of vitamin C supplementation reduces amyloid plaque burden and ameliorates pathological changes in the brain of 5XFAD mice. *Cell Death Dis* 2014; 5(2): e1083.  
<http://dx.doi.org/10.1038/cddis.2014.26> PMID: 24577081
- [37] Cha J, Roomi MW, Kalinovsky T, Niedzwiecki A, Rath M. Lipoprotein(a) and vitamin C impair development of breast cancer tumors in Lp(a)+; Gulo-/- mice. *Int J Oncol* 2016; 49(3): 895-902.  
 38. Naidu KA. Vitamin C in human health and disease is still a mystery? An overview. *Nutr J* 2003; 2: 7.
- [38] Naidu KA. Vitamin C in human health and disease is still a mystery? An overview. *Nutr J* 2003; 2: 7.  
<http://dx.doi.org/10.1186/1475-2891-2-7> PMID: 14498993
- [39] Hodges RE, Hood J, Canham JE, Sauberlich HE, Baker EM. Clinical manifestations of ascorbic acid deficiency in man. *Am J Clin Nutr* 1971; 24(4): 432-43.  
<http://dx.doi.org/10.1093/ajcn/24.4.432> PMID: 5090631
- [40] Aumailley L, Warren A, Garand C, *et al.* Vitamin C modulates the metabolic and cytokine profiles, alleviates hepatic endoplasmic reticulum stress, and increases the life span of Gulo-/- mice. *Aging (Albany NY)* 2016; 8(3): 458-83.  
<http://dx.doi.org/10.18632/aging.100902> PMID: 26922388
- [41] Kuo SM, MacLean ME, McCormick K, Wilson JX. Gender and sodium-ascorbate transporter isoforms determine ascorbate concentrations in mice. *J Nutr* 2004; 134(9): 2216-21.  
<http://dx.doi.org/10.1093/jn/134.9.2216> PMID: 15333707
- [42] Tsukaguchi H, Tokui T, Mackenzie B, *et al.* A family of mammalian Na<sup>+</sup>-dependent L-ascorbic acid transporters. *Nature* 1999; 399(6731): 70-5.  
<http://dx.doi.org/10.1038/19986> PMID: 10331392
- [43] Tveden-Nyborg P, Hasselholt S, Miyashita N, Moos T, Poulsen HE, Lykkesfeldt J. Chronic vitamin C deficiency does not accelerate oxidative stress in ageing brains of guinea pigs. *Basic Clin Pharmacol Toxicol* 2012; 110(6): 524-9.  
<http://dx.doi.org/10.1111/j.1742-7843.2011.00852.x> PMID: 22212866
- [44] Abad-Rodriguez J, Ledesma MD, Craessaerts K, *et al.* Neuronal membrane cholesterol loss enhances amyloid peptide generation. *J Cell Biol* 2004; 167(5): 953-60.  
<http://dx.doi.org/10.1083/jcb.200404149> PMID: 15583033
- [45] Pierrot N, Tyteca D, D'auria L, *et al.* Amyloid precursor protein controls cholesterol turnover needed for neuronal activity. *EMBO Mol Med* 2013; 5(4): 608-25.  
<http://dx.doi.org/10.1002/emmm.201202215> PMID: 23554170
- [46] Svennerholm L, Boström K, Helander CG, Jungbjer B. Membrane lipids in the aging human brain. *J Neurochem* 1991; 56(6): 2051-9.  
<http://dx.doi.org/10.1111/j.1471-4159.1991.tb03466.x> PMID: 2027013
- [47] Izzotti A, Cartiglia C, Taningher M, De Flora S, Balansky R. Age-related increases of 8-hydroxy-2'-deoxyguanosine and DNA-protein crosslinks in mouse organs. *Mutat Res* 1999; 446(2): 215-23.  
[http://dx.doi.org/10.1016/S1383-5718\(99\)00189-8](http://dx.doi.org/10.1016/S1383-5718(99)00189-8) PMID: 10635344
- [48] Tian L, Cai Q, Wei H. Alterations of antioxidant enzymes and oxidative damage to macromolecules in different organs of rats during aging. *Free Radic Biol Med* 1998; 24(9): 1477-84.  
[http://dx.doi.org/10.1016/S0891-5849\(98\)00025-2](http://dx.doi.org/10.1016/S0891-5849(98)00025-2) PMID: 9641266
- [49] Mecocci P, MacGarvey U, Kaufman AE, *et al.* Oxidative damage to mitochondrial DNA shows marked age-dependent increases in human brain. *Ann Neurol* 1993; 34(4): 609-16.  
<http://dx.doi.org/10.1002/ana.410340416> PMID: 8215249
- [50] HariPriya D, Sangeetha P, Kanchana A, Balu M, Panneerselvam C. Modulation of age-associated oxidative DNA damage in rat brain cerebral cortex, striatum and hippocampus by L-carnitine. *Exp Gerontol* 2005; 40(3): 129-35.  
<http://dx.doi.org/10.1016/j.exger.2004.10.006> PMID: 15763389
- [51] Gemma C, Vila J, Bachstetter A, Bachstetter A, Bickford PC. Oxidative stress and the aging brain: From theory to prevention. In: *Brain Aging: Models, Methods, and Mechanisms* Frontiers in Neuroscience. Boca Raton, FL: CRC Press/Taylor & Francis 2007.  
<http://dx.doi.org/10.1201/9781420005523-15>
- [52] Lovell MA, Markesbery WR. Oxidative DNA damage in mild cognitive impairment and late-stage Alzheimer's disease. *Nucleic Acids Res* 2007; 35(22): 7497-504.  
<http://dx.doi.org/10.1093/nar/gkm821> PMID: 17947327
- [53] Frazer KA, Narla G, Zhang JL, Rubin EM. The apolipoprotein(a) gene is regulated by sex hormones and acute-phase inducers in YAC transgenic mice. *Nat Genet* 1995; 9(4): 424-31.  
<http://dx.doi.org/10.1038/ng0495-424> PMID: 7795650
- [54] Acquati F, Hammer R, Ercoli B, *et al.* Transgenic mice expressing a human apolipoprotein[a] allele. *J Lipid Res* 1999; 40(6): 994-1006.  
[http://dx.doi.org/10.1016/S0022-2275\(20\)33503-3](http://dx.doi.org/10.1016/S0022-2275(20)33503-3) PMID: 10357831
- [55] Suk Danik J, Rifai N, Buring JE, Ridker PM. Lipoprotein(a), hormone replacement therapy, and risk of future cardiovascular events. *J Am Coll Cardiol* 2008; 52(2): 124-31.  
<http://dx.doi.org/10.1016/j.jacc.2008.04.009> PMID: 18598891
- [56] Kurl S, Tuomainen TP, Laukkanen JA, *et al.* Plasma vitamin C modifies the association between hypertension and risk of stroke. *Stroke* 2002; 33(6): 1568-73.  
<http://dx.doi.org/10.1161/01.STR.0000017220.78722.D7> PMID: 12052992
- [57] Mahmoodian F, Peterkofsky B. Vitamin C deficiency in guinea pigs differentially affects the expression of type IV collagen, laminin, and elastin in blood vessels. *J Nutr* 1999; 129(1): 83-91.  
<http://dx.doi.org/10.1093/jn/129.1.83> PMID: 9915880
- [58] Obata F, Narita K. Hypercholesterolemia negatively influences morphology and molecular markers of epithelial cells within the choroid plexus in rabbits. *Fluids Barriers CNS* 2020; 17(1): 13.  
<http://dx.doi.org/10.1186/s12987-020-0175-0> PMID: 32019573